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Lee et al.

(54) MICROORGANISM PRODUCING 4-HYDROXYBUTYRATE AND A METHOD FOR PRODUCING 4-HYDROXYBUTYRATE IN ANAEROBIC CONDITION USING THE SAME

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(58) Field of Classification Search

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(57) ABSTRACT

A genetically modified microorganism comprising a polynucleotide encoding α -ketoglutarate synthase or a mutant thereof, and a polynucleotide encoding pyruvate carboxylase or a mutant thereof; wherein the genetically modified microorganism has decreased malate quinone oxidoreductase activity and/or decreased phosphoenolpyruvate carboxykinase activity compared to an unmodified microorganism of the same type, and wherein the genetically modified microorganism produces 4-hydroxybutyrate.

19 Claims, 4 Drawing Sheets

FIG. 1

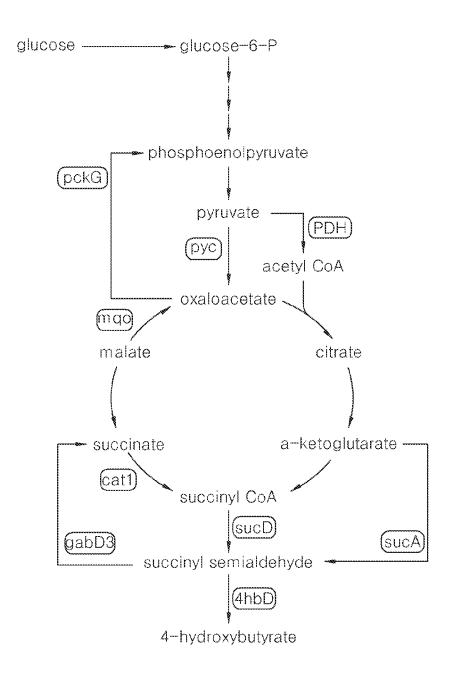


FIG. 2

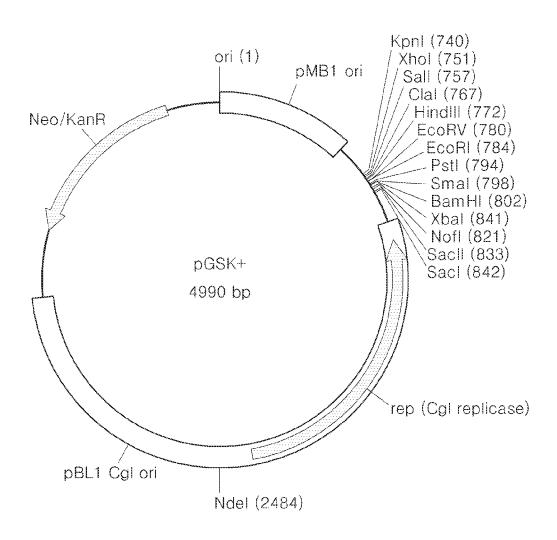


FIG. 3

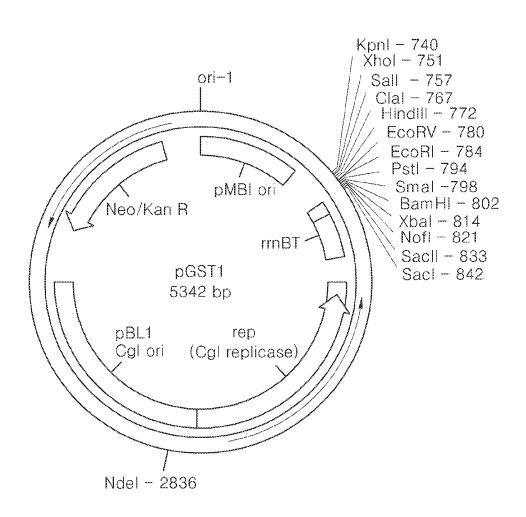


FIG. 4

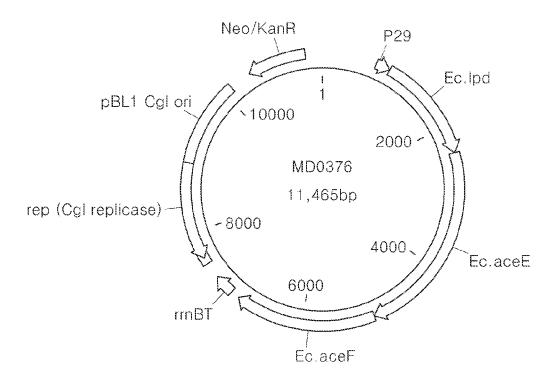
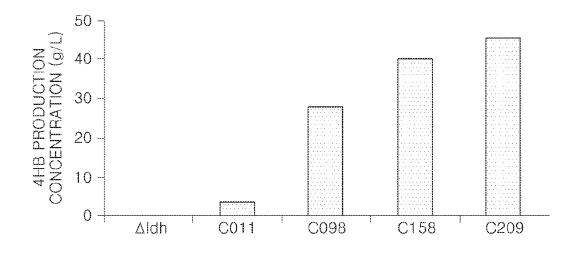


FIG. 5



MICROORGANISM PRODUCING 4-HYDROXYBUTYRATE AND A METHOD FOR PRODUCING 4-HYDROXYBUTYRATE IN ANAEROBIC CONDITION USING THE **SAME**

CROSS-REFERENCE TO RELATED APPLICATIONS

This application claims the benefit of Korean Patent Application No. 10-2013-0100568, filed on Aug. 23, 2013, in the Korean Intellectual Property Office, the entire disclosure of which is hereby incorporated by reference.

INCORPORATION BY REFERENCE OF ELECTRONICALLY SUBMITTED MATERIALS

Incorporated by reference in its entirety herein is a computer-readable nucleotide/amino acid sequence listing sub- 20 mitted concurrently herewith and identified as follows: One 149,196 Byte ASCII (Text) file named "718145 ST25-Revised.TXT" created on Nov. 3, 2014.

BACKGROUND

1. Field

The present disclosure relates to a genetically modified microorganism that produces 4-hydroxybutyrate and a method of producing 4-hydroxybutyrate using the geneti- 30 cally modified microorganism.

2. Description of the Related Art

Biodegradable polymers have been suggested as an alternative to the synthetic polymers which account for a great part of severe environmental pollution. Accordingly, various 35 cally engineered strain. biodegradable polymers have been developed. One such biodegradable polymer is poly-β-hydroxybutyrate, which is a biodegradable polymer accumulated in a nutritionally imbalanced state by various microorganisms and has excelpiezoelectricity, and biocompatibility. 4-hydroxybutyrate (4HB), which is one of the various types of poly-β-hydroxybutyrate, is a representative polyhydroxyalkanoate (PHA). 4HB is a substance produced as a white powder in a small quantity from wine, beef, and fruit. Many studies are con- 45 ducted with regard to 4HB as a biodegradable plastic material as 4HB shows a wide range of physical properties from crystalline plastic to highly elastic rubber as 4HB is similar to polyester. 4HB for medical use is generally produced by fermentation. Although a method of producing 50 1,4-butandiol (1,4-BDO) from 4HB is known, it has not been commercialized yet.

4HB has been used as a starting material in producing other C4-chemicals such as 1,4-BDO and γ-butyrolactone (GBL) by methods using a microorganism. 4HB may easily 55 be converted to various other C4-chemicals such as 1,4-BDO, GBL, and tetrahydrofuran (THF). These various chemicals are used in the chemical industries as polymer, solvent, and fine chemical intermediates.

Most C4-chemicals that are currently synthesized are 60 derived from 1,4-butandiol or maleic anhydride, but the chemical production process needs to be improved or replaced by a newly developed process as production costs are increasing due to rising oil prices. A biological process for producing C4-chemicals is suggested as an alternative to 65 the chemical process, but the yield of 4HB production using conventional microorganisms is low. Thus, there is a need

2

for a mutant microorganism capable of producing 4HB, and a biological method of producing 4HB using the mutant microorganism.

SUMMARY

Provided is a genetically modified (i.e. engineered) microorganism that produces 4-hydroxybutyrate (4HB), and a method of increasing 4HB production by using the prepared strain. The genetically modified microorganism comprises a polynucleotide encoding α-ketoglutarate synthase or a mutant thereof, and a polynucleotide encoding pyruvate carboxylase or a mutant thereof; and has decreased malate quinone oxidoreductase activity and/or decreased phospho-15 enolpyruvate carboxykinase activity compared to an unmodified microorganism of the same type.

BRIEF DESCRIPTION OF THE DRAWINGS

These and/or other aspects will become apparent and more readily appreciated from the following description of the embodiments, taken in conjunction with the accompanying drawings of which:

FIG. 1 is a flow chart displaying genes for alteration, 25 deletion, and introduction in metabolic pathways and glycolysis pathways for 4HB production in a microorganism;

FIG. 2 is a map of the expression vector pGSK+ used in preparing a genetically modified microorganism;

FIG. 3 is a map of the expression vector pGST1 used in preparing a genetically modified microorganism;

FIG. 4 is a map of the expression vector MD0376 used in preparing a genetically modified microorganism; and

FIG. 5 is a graph comparing 4HB production concentration between the basic Corynebacteria strain and a geneti-

DETAILED DESCRIPTION

Reference will now be made in detail to embodiments, lent properties such as biodegradability, moisture resistance, 40 examples of which are illustrated in the accompanying drawings, wherein like reference numerals refer to the like elements throughout. In this regard, the present embodiments may have different forms and should not be construed as being limited to the descriptions set forth herein. Accordingly, the embodiments are merely described below, by referring to the figures, to explain aspects. As used herein, the term "and/or" includes any and all combinations of one or more of the associated listed items.

> Provided is a genetically modified microorganism (i.e., a strain) that produces 4-hydroxybutyrate (4HB).

> An embodiment of invention provides a genetically modified microorganism that produces 4HB, wherein the activity of malate quinone oxidoreductase and/or the activity of phosphoenolpyruvate carboxykinase is eliminated or decreased compared to an unmodified microorganism of the same type, and wherein the genetically modified microorganism comprises a polynucleotide encoding α-ketoglutarate synthase or a mutant thereof, and/or a polynucleotide encoding pyruvate carboxylase or a mutant thereof.

> The term "unmodified microorganism of the same type" means a reference microorganism that is compared to a genetically modified microorganism comprising a subject modification. The reference microorganism refers to a wildtype microorganism or a parental microorganism. The parental microorganism refers to a microorganism that has not undergone the subject modification that the genetically modified microorganism has undergone and is genetically

identical to the genetically modified microorganism except for the modification, and thus serves as a reference microorganism for the modification.

In addition, the genetically modified microorganism may be a microorganism wherein the activity of succinate semialdehyde dehydrogenase is eliminated or decreased.

In addition, the genetically modified microorganism may further include a polynucleotide encoding pyruvate dehydrogenase or a mutant thereof.

In addition, the genetically modified microorganism may 10 further include a polynucleotide encoding formate dehydrogenase or a mutant thereof.

The strain may be a strain selected from the group consisting of lumen bacteria, Corynebacterium genus, Brevibacterium genus, and Escherichia coli. The strain may 15 be Corynebacterium glutamicum. In particular, Corynebacterium glutamicum may be cultured in a wide range of culture conditions and at a high growth rate. In addition, Corynebacterium glutamicum is non-pathogenic and harmless to environment, as they do not produce a spore. In 20 particular, Corynebacterium glutamicum is highly available in industries as it may be cultured to a concentration four times higher than that of Escherichia coli.

The strain capable of producing 4HB may be a strain or decreased compared to an unmodified microorganism of the same type. The strain capable of producing 4HB may include succinyl-CoA:coenzyme A transferase or a mutant thereof, coenzyme-dependent succinate semialdehyde dehydrogenase or a mutant thereof, and 4-hydroxybutyrate dehydrogenase or a mutant thereof.

Lactate dehydrogenase is an enzyme that catalyzes the conversion of pyruvate to lactate. The lactate dehydrogenase may include lactate dehydrogenase (Ldh), lactate dehydrogenase A (LdhA), lactate dehydrogenase B (LdhB), and 35 lactate dehydrogenase C (LdhC). The activity of the lactate dehydrogenase may be eliminated or decreased in a genetically modified microorganism. The lactate dehydrogenase may an enzyme classified as EC.1.1.1.27. The lactate dehydrogenase may be referred to as LdhA. The genetically 40 modified microorganism may be a microorganism wherein a gene encoding lactate dehydrogenase is inactivated or attenuated. The mutant of lactate dehydrogenase may be an enzyme having catalytic activity the same as that of lactate dehydrogenase and sequence identity of 80% or higher with 45 amino acid sequence of a wild type lactate dehydrogenase. The mutant may be an enzyme having catalytic activity the same as that of lactate dehydrogenase and sequence identity of 85% or higher, 90% or higher, 95% or higher, or 99% or higher with amino acid sequence of a wild type lactate 50 dehydrogenase.

The succinyl-CoA:coenzyme A transferase may be referred to as Cat1. The enzyme catalyzes the conversion of succinate to succinyl-CoA. The succinyl-CoA:coenzyme A transferase may be an enzyme classified as EC.2.8.3. The 55 enzyme may be derived from Corynebacterium glutamicum or Clostridium kluyveri. The succinyl-CoA:coenzyme A transferase may have an amino acid sequence of SEQ ID NO: 1. A polynucleotide encoding the succinyl-CoA:coenzyme A transferase may have a nucleic acid of SEQ ID NO: 60 2. The mutant of the succinyl-CoA:coenzyme A transferase may be an enzyme having catalytic activity the same as that of the succinyl-CoA:coenzyme A transferase and sequence identity of 80% or higher with amino acid sequence of a wild type the succinyl-CoA:coenzyme A transferase. The mutant 65 may be an enzyme having catalytic activity the same as that of the succinyl-CoA:coenzyme A transferase and sequence

identity of 85% or higher, 90% or higher, 95% or higher, or 99% or higher with amino acid sequence of a wild type the succinyl-CoA:coenzyme A transferase.

The CoA-dependent succinate semialdehyde dehydrogenase may be referred to as SucD. The enzyme catalyzes the conversion of succinyl-CoA to succinyl semialdehyde. A polynucleotide encoding the CoA-dependent succinate semialdehyde dehydrogenase (SEQ ID NO: 3) may be derived from a Corynebacterium glutamicum or Porphyromonas gingivalis. The polynucleotide encoding the CoAdependent succinate semialdehyde dehydrogenase may have a nucleic acid of SEQ ID NO: 4. The mutant of CoAdependent succinate semialdehyde dehydrogenase may be an enzyme having catalytic activity the same as that of CoA-dependent succinate semialdehyde dehydrogenase and sequence identity of 80% or higher with amino acid sequence of a wild type succinate semialdehyde dehydrogenase. The mutant may be an enzyme having catalytic activity the same as that of CoA-dependent succinate semialdehyde dehydrogenase and sequence identity of 85% or higher, 90% or higher, 95% or higher, or 99% or higher with amino acid sequence of a CoA-dependent succinate semialdehyde dehydrogenase.

The 4-hydroxybutyrate dehydrogenase may be referred to wherein the activity of lactate dehydrogenase is eliminated 25 as 4Hbd. The enzyme catalyzes the conversion of succinyl semialdehyde to 4HB. The enzyme may be derived from a Corynebacterium glutamicum or Porphyromonas gingivalis. The 4-hydroxybutyrate dehydrogenase may have an amino acid sequence of SEQ ID NO: 5. A polynucleotide encoding the 4-hydroxybutyrate dehydrogenase may have a nucleic acid of SEQ ID NO: 6. The mutant of 4-hydroxybutyrate dehydrogenase may be an enzyme having catalytic activity the same as that of 4-hydroxybutyrate dehydrogenase and sequence identity of 80% or higher with amino acid sequence of a wild type 4-hydroxybutyrate dehydrogenase. The mutant may be an enzyme having catalytic activity the same as that of 4-hydroxybutyrate dehydrogenase and sequence identity of 85% or higher, 90% or higher, 95% or higher, or 99% or higher with amino acid sequence of a wild type 4-hydroxybutyrate dehydrogenase.

> A gene may be additionally introduced into or eliminated from a strain capable of producing 4HB. For example, activity of malate quinone oxidoreductase may be eliminated or decreased in the strain compared to an unmodified microorganism of the same type. Malate quinone oxidoreductase may be referred to as Mgo. The enzyme catalyzes the conversion of converting malate to oxaloacetate. The malate quinone oxidoreductase may have an amino acid sequence of SEQ ID NO: 17. Gene addition, substitution, or deletion may be performed in a polynucleotide encoding malate quinone oxidoreductase in order to decrease activity of malate quinone oxidoreductase. The polynucleotide encoding malate quinone oxidoreductase may be partly or totally deleted by homologous recombination. The polynucleotide encoding malate quinone oxidoreductase may have a nucleic acid sequence of SEQ ID NO: 18.

> Activity of phosphoenolpyruvate carboxykinase may be eliminated or decreased in the strain compared to an unmodified microorganism of the same type. Phosphoenolpyruvate carboxykinase may be referred to as PckG. The enzyme catalyzes the conversion of converting oxaloacetate to phosphoenolpyruvate. The phosphoenolpyruvate carboxykinase may have an amino acid sequence of SEQ ID NO: 19. Gene addition, substitution, or deletion may be performed in a polynucleotide encoding phosphoenolpyruvate carboxykinase in order to decrease activity of phosphoenolpyruvate carboxykinase. The polynucleotide encod-

ing phosphoenolpyruvate carboxykinase may be partly or totally deleted by homologous recombination. The polynucleotide encoding phosphoenolpyruvate carboxykinase may have a nucleic acid sequence of SEQ ID NO: 20.

The microorganism may include a polynucleotide encod- 5 ing α-ketoglutarate synthase or a mutant thereof. α-ketoglutarate synthase may be referred to as SucA. The enzyme catalyzes the conversion of ketoglutarate to succinyl semialdehyde. The enzyme may be derived from Corynebacterium glutamicum or Mycobacterium bovis. The α-ketoglutarate synthase may have an amino acid sequence of SEQ ID NO: 7. A polynucleotide encoding the α -ketoglutarate synthase may have a nucleic acid of SEQ ID NO: 8. The mutant of α-ketoglutarate synthase is an enzyme having catalytic activity the same as that of \alpha-ketoglutarate synthase and 15 sequence identity of 80% or higher with amino acid sequence of a wild type α -ketoglutarate synthase. The mutant may be an enzyme having catalytic activity the same as that of α -ketoglutarate synthase and sequence identity of 85% or higher, 90% or higher, 95% or higher, or 99% or 20 higher with amino acid sequence of a wild type α -ketoglutarate synthase.

The microorganism may include a polynucleotide encoding pyruvate carboxylase or a mutant thereof. Pyruvate carboxylase may be referred to as Pyc. The enzyme cata- 25 lyzes the conversion of pyruvate to oxaloacetate. The enzyme may be derived from Corynebacterium glutamicum or Escherichia coli. The pyruvate carboxylase may have an amino acid sequence of SEQ ID NO: 9. The mutant of pyruvate carboxylase is an enzyme having catalytic activity 30 the same as that of pyruvate carboxylase and sequence identity of 80% or higher with amino acid sequence of a wild type pyruvate carboxylase. The mutant may be an enzyme having catalytic activity the same as that of pyruvate carboxylase and sequence identity of 85% or higher, 90% or 35 higher, 95% or higher, or 99% or higher with amino acid sequence of a wild type pyruvate carboxylase. The mutant of pyruvate carboxylase may be formed by substituting the 458th amino acid of a wild type pyruvate carboxylase. The 458th amino acid of a wild type pyruvate carboxylase may 40 be proline. The 458th amino acid of a wild type pyruvate carboxylase may be substituted with serine (SEQ ID NO: 10). The polynucleotide encoding pyruvate carboxylase may have a nucleic acid sequence of SEQ ID NO: 11.

Activity of succinate semialdehyde dehydrogenase may 45 be eliminated or decreased in the strain. Succinate semialdehyde dehydrogenase may be referred to as SSADH. The enzyme catalyzes the conversion of succinyl semialdehyde to succinate. The succinate semialdehyde dehydrogenase may have an amino acid sequence of SEQ ID NO:21. Gene 50 addition, substitution, or deletion may be performed in NCgl0049, NCgl0463, or NCgl2619 gene in order to decrease activity of succinate semialdehyde dehydrogenase. The NCgl0049 polynucleotide may be partly or totally deleted by homologous recombination. The NCgl0049 polynucleotide may have a nucleic acid sequence of SEQ ID NO: 22. The NCgl0463 polynucleotide may have a nucleic acid sequence of SEQ ID NO: 24.

The microorganism may include a polynucleotide encoding pyruvate dehydrogenase. The pyruvate dehydrogenase may be referred to as "pyruvate dehydrogenase complex" and also Pdh. The pyruvate dehydrogenase catalyzes the conversion of to acetyl CoA. The pyruvate dehydrogenase complex includes pyruvate dehydrogenase (E1), dihydrolipoyl transacetylase (E2), and dihydrolipoyl dehydrogenase (E3). In the pyruvate dehydrogenase, E1 is also referred to

6

as AceE, E2 is referred to as AceF, and E3 is referred to as lpd or lpdA, depending on microorganisms.

A polynucleotide encoding the pyruvate dehydrogenase includes lpd, aceE, and aceF genes. The genes may be derived from *Corynebacterium glutamicum* or *Escherichia coli*. The lpd gene may be a polynucleotide having a nucleic acid sequence of SEQ ID NO: 12. The aceE gene may be a polynucleotide having a nucleic acid sequence of SEQ ID NO: 13. The aceF gene may be a polynucleotide having a nucleic acid sequence of SEQ ID NO: 14.

The microorganism may include a polynucleotide encoding formate dehydrogenase or a mutant thereof. Formate dehydrogenase may be referred to as Fdh. The enzyme catalyzes the conversion of formate to bicarbonate. The enzyme may be derived from *Corynebacterium glutamicum* or *Mycobacterium vaccae*. The formate dehydrogenase may have an amino acid sequence of SEQ ID NO: 15. The polynucleotide encoding formate dehydrogenase may have a nucleic acid sequence of SEQ ID NO: 16.

The polynucleotide encoding an enzyme may be introduced to a strain as it is inserted into a vector. The polynucleotide may be operably linked with a regulatory sequence. A regulatory sequence, which is a sequence regulating expression of the polynucleotide, may include a promoter, a terminator, or an enhancer.

The term "vector" refers to a DNA product including a DNA sequence operably linked with an appropriate regulation sequence capable of expressing DNA in an appropriate host cell. The vector may be a plasmid vector, a bacteriophage vector, or a cosmid vector. To operate as an expression vector, a vector may include a replication origin, a promoter, a multi-cloning site (MCS), a selection marker or a combination thereof. A replication origin gives a function to a plasmid to replicate itself independently of hose cell chromosome. A promoter operates in transcription process of an inserted foreign gene. An MCS enables a foreign gene to be inserted through various restriction enzyme sites. A selection marker verifies whether a vector has been properly introduced to a host cell. A selection includes an antibioticresistant gene generally used in the art. For example, a selection marker may include a gene resistant to ampicillin, gentamycin, carbenicillin, chloramphenicol, streptomycin, kanamycin, geneticin, neomycin or tetracycline. Considering the cost, ampicillin or gentamycin-resistant gene may be used.

When a vector of an embodiments uses a prokaryotic cell as host cell, a strong promoter, for example, lamda-PL promoter, trp promoter, lac promoter or T7 promoter, is included in the vector. If a vector uses a eukaryotic cell as host cell, the vector may include a promoter derived from genome of a mammal (metallothionin promoter, e.g.) or a promoter derived from a mammal virus (adenovirus late promoter, vaccinia virus 7.5K promoter, SV40 promoter, cytomegalovirus promoter or tk promoter of HSV promoter, e.g.). The promoter may be a lamda-PL promoter, trp promoter, lac promoter or T7 promoter. In this manner, a promoter is operably linked with a sequence encoding a gene.

The promoter is operably linked with a sequence encoding a gene. The term "operably linked" herein means a functional bond between a nucleic acid expression regulatory sequence (e.g. promoter, signal sequence or array at transcription regulation factor binding site, a terminator, or an enhancer) and another nucleic acid sequence. Through the functional bond, the regulatory sequence may control transcription and/or translation of a nucleotide encoding the gene.

The term "transformation" herein refers to introducing a gene to a host cell so that the gene may be expressed in the microorganism by methods known in the art (e.g., heat shock and electroporation). A transformed gene, only if the gene may be expressed in the host cell, may be any gene 5 whether the gene is inserted into a chromosome of the host cell or the gene exists outside a chromosome. The gene, which is a polynucleotide capable of encoding a polypeptide, may be DNA or RNA. The introduction of the gene may be any type of introduction, only if the gene may be 10 introduced into and expressed in the host cell. For example, the gene may be introduced into a host cell by an introduction in the form of an expression cassette, which is a polynucleotide structure including all factors related to the expression of the gene by itself. The expression cassette 15 usually includes a promoter, a transcription termination signal, a ribosome binding site, and a translation termination signals operably linked with the gene. The expression cassette may be an expression vector capable of self-replication. In addition, the gene may be introduced as itself or in the 20 form of a polynucleotide structure to a host cell and then be operably linked with a sequence related to an expression in the host cell.

Attenuation of activity of the enzyme maybe performed altered so that enzyme activity may be weakened or deleted, by substituting a promoter of the gene with a promoter weaker than an endogenous promoter, or by deleting the gene from a chromosome. A gene encoding an enzyme may be deleted from genome of a microorganism by homologous 30 recombination.

Another aspect relates to a method of producing C4-chemicals by culturing the genetically modified microorganism in a cell culture medium, whereby the microorganism produces a C4-chemical; and recovering the 35 C4-chemical from the culture solution. The C4-chemicals may include succinic acid, succinate, fumaric acid, malic acid, or a C4 chemical derived therefrom. For example, production of C4-chemicals included in TCA cycle or substances derived therefrom may be increased by culturing the 40 microorganism of an aspect. In addition, the substances derived from succinate may be 4-HB, 1,4-BDO, γ-butyrolactone (GBL) or C4 chemicals derived therefrom but are not limited thereto.

The culturing may be performed under an appropriate 45 culture medium composition and culture conditions known in this art. The culture medium composition and culture conditions may be conveniently adjusted according to the selected microorganism. The culturing method may include batch culturing, continuous culturing, fed-batch culturing or 50 a combination thereof. The fed-batch culturing may use a culture medium having glucose 50 g/L, corn steep liquor 10 g/L, (NH₄)₂SO₄ 45 g/L, UREA 4.5 g/L, KH₂PO₄ 0.5 g/L, MgSO₄/7H₂O 0.5 g/L, FeSO₄/7H₂O (10 g/L) stock 1 mL, MnSO₄/4H₂O (10 g/L) stock 1 mL, beta-alanin (5 g/L) stock 55 1 mL, nicotinic acid (5 g/L) stock 1 mL, thiamine-HCl (5 g/L) stock 1 mL, and D-biotin (0.3 g/L) stock 1 mL. The culture condition may comprise a combination of aerobic and anaerobic conditions. For example, the genetically modified microorganism may be cultured under aerobic 60 condition for 20 hr and subsequently, under anaerobic condition for 80 hr.

The culture medium may include various carbon sources, nitrogen sources, and trace elements. The carbon source may include a carbohydrate such as glucose, sucrose, lactose, 65 fructose, maltose, starch, and cellulose, a lipid such as soybean oil, sunflower oil, castor oil, and coconut oil, a fatty

acid such as palmitic acid, stearic acid, and linoleic acid, an organic acid such as acetic acid or a combination thereof. The culturing may be performed by using glucose as a carbon source. The nitrogen source may include an organic nitrogen source such as peptone, yeast extract, meat extract, malt extract, corn steep liquid, and soybean, an inorganic nitrogen source such as urea, ammonium sulfate, ammonium chloride, ammonium phosphate, ammonium carbonate, and ammonium nitrate or a combination thereof. The culture medium may include as a phosphorous source, for example, potassium dihydrogen phosphate, dipotassium phosphate, a sodium-containing salt corresponding to potassium dihydrogen phosphate, and dipotassium phosphate, and a metal salt such as magnesium sulfate and iron sulfate. The culture medium or an individual component may be added to the culture in a batch mode or a continuous mode.

In addition, pH of the culture may be adjusted during the culturing by adding a compound such as ammonium hydroxide, potassium hydroxide, ammonia, phosphoric acid or sulfuric acid to the culture in an appropriate mode. In addition, bubble formation may be repressed by using an endoplasmic reticulum such as fatty acid polyglycol ester.

The microorganism may be cultured under anaerobic by substituting an endogenous gene with a gene which is 25 conditions. The term "anaerobic conditions" herein refers to a state wherein oxygen content is lower than that of normal atmospheric state. Anaerobic conditions may be formed, for example, by supplying carbon dioxide or nitrogen at a flow rate range from about 0.1 vvm (Volume per Volume per Minute) to about 0.4 vvm, from about 0.2 vvm to about 0.3 vvm or at a flow rate of 0.25 vvm. In addition, anaerobic conditions may be formed by setting an aeration rate in the range from about 0 vvm and to 0.4 vvm, from about 0.1 vvm to about 0.3 vvm or from 0.15 vvm to about 0.25 vvm.

> The method of producing C4-chemicals includes recovering of the produced C4-chemicals from the culture. The produced C4-chemicals may be succinic acid, succinate, fumaric acid, malic acid or a C4-chemical derived therefrom. According to one embodiment, the produced C4-chemicals may be 4-HB, 1,4-BDO, GBL or a C4-chemical derived therefrom. For example, the recovery of 4-HB may be performed by using known separation and purification methods. The recovery may be performed by centrifugation, ion exchange chromatography, filtration, precipitation or a combination thereof. Recovery of C4-chemicals, for example, recovery of succinic acid, 4HB, or GBL may be performed by a method known in this art including filtration of culture solution.

> In addition, the method of producing C4-chemicals may be used to produce other various organic compounds by converting the C4-chemicals to other organic chemicals. A substrate structurally related to 4-HB may be synthesized by chemically converting the 4-HB yielded in the method described above. According to one embodiment, gamma butyrolactone (GBL) may be yielded by reacting 4-HB at about 100° C. to 200° C. in the presence of a strong acid and then distilling the reactant. The yielded GBL may be converted to N-methyl pyrrolidone (NMP) by amination using an aminating agent, for example, methylamine. In addition, the yielded GBL may be selectively converted to tetrahydrofuran (THF), 1,4-BDO or butanol by hydrogenation using a metal-containing catalyst, for example, Ru or Pd.

> The poly-4-hydroxybutyrate may be yielded by biologically converting the produced 4-HB. The biological conversion may be achieved by polyhydroxyalkanoate synthase, 4-HB-CoA:coenzyme A transferase or a combination thereof.

As described above, according to the one or more of the above embodiments, yield of 4HB production was improved by performing additional genetic engineering in a genetically modified microorganism that produces 4HB. In particular, activity of enzymes involved in various metabolic pathways was altered, and a genetically modified microorganism capable of producing 4HB at the optimal yield was developed by a combination of various genetic mutations. As the genetically modified microorganism having increased 4HB productivity may produce 4HB at a high yield by anaerobic fermentation, the strain may be very useful in industries.

A genetically modified microorganism capable of producing 4HB at a high efficiency in anaerobic conditions may be effectively used in 4HB production. As 4HB is a compound highly useful in industries, 4HB productivity increase in the strain may increase utility of 4HB by reducing unit price of 4HB production. Therefore, the transformed microorganism may be very useful in industries.

It should be understood that the exemplary embodiments described therein should be considered in a descriptive sense only and not for purposes of limitation. Descriptions of ²⁵ features or aspects within each embodiment should typically be considered as available for other similar features or aspects in other embodiments.

Example 1

Preparation of Transformed Strain

- 1.1 Preparation of CGL (Δldh) Strain
- (1) Preparation of Replacement Vector

To prevent excessive lactate accumulation in natural *Corynebacterium* during anaerobic fermentation, a gene (SEQ ID NO: 42) expressing L-lactate dehydrogenase (NCgl2810) in the strain was eliminated. An open reading frame (ORF) region of 230 bp in the gene was eliminated by gene substitution. Sequence of the eliminated region is shown in SEQ ID NO: 29.

An upstream region corresponding to first 200 bp of ldhA 45 gene was amplified by PCR using genome DNA of CGL ATCC 13032 as a template and using ldhA up 5' (SEQ ID NO:25) and ldhA_up_3' (SEQ ID NO: 26) primer sets. A downstream region corresponding to last 200 bp of ldhA 50 gene was amplified by PCR using primer sets ldhA_down_5' (SEQ ID NO:27) and ldhA_down_3' (SEQ ID NO:28). The PCR amplification was performed by repeating, 30 times, a cycle including a denaturation step at 95° C. for 30 seconds, an annealing step at 55° C. for 30 seconds, and an extension step at 72° C. for 30 seconds. All the PCR amplifications hereinafter were performed under the same conditions. Afterward, flanking regions upstream and downstream from ldhA gene was overlapped with an extended region of an 60 oligonucleotide and linked with each other by using 16 bp In-Fusion® HD Cloning Kit (cat no. 639691 manufactured by Clontech). The linked regions upstream and downstream from the ldhA product were ligated with pK19mobsacB 60 (obtained from ATCC 87098) at BamHI and SalI restriction enzyme sites. As a result, pK19mobsacB-Δldh was obtained.

10

(2) Preparation of CGL (Δldh) Strain

The obtained plasmid, pK19mobsacB-ΔldhA, was used to delete a corresponding gene in CGL by homologous recombination according to Schafer et al. (1994). The pK19mobsacB-Δldh vector was introduced to CGL ATCC13032 bp electroporation. The strain wherein the vector was introduced was cultured at 30° C. by streaking the strain on lactobacillus selection (LBHIS) culture medium including kanamycin 25 μg/ml. The LBHIS culture medium includes brain-heart infusion broth 18.5 g/L, 0.5 M sorbitol, 5 g/L bacto-tryptone, 2.5 g/L bacto-yeast extract, 5 g/L NaCl, and 18 g/L bacto-agar. Hereinafter, composition of LBHIS medium is the same. The colony was streaked on LB-sucrose culture medium and cultured at 30° C. Then, only the colonies wherein double crossing-over occurred were selected. Gene deletion was verified by PCR using primer sets (SEQ ID NO: 25 and SEQ ID NO: 28). The obtained strain was named as $CGL(\Delta ldh)$.

TABLE 1

SEQ ID NO	Sequence
25	5'-GCAGGCATGCAAGCTTCTAGTCTGGGGAGCGAAACC-3'
26	5'-GAGCTCAGTCATCGACGCCACGAGGAAGATG-3'
27	5'-TGACTGACTGAGCTCCTGGACAAAGACCCAGAGCT-3'
28	5'-GGCCAGTGCCAAGCTTTTGCGGGCACCAACGTAATG-3'

1.2 Preparation of Strain Capable of Producing 4HB

The CGL(Δ ldh) strain prepared in Example 1.1 was genetically engineered further so that the strain may become capable of producing 4HB. In order to attenuate NCgl0049 gene, a polynucleotide encoding succinyl-CoA:coenzyme A transferase, CoA-dependent succinate semialdehyde dehydrogenase, and 4-hydroxybutyrate dehydrogenase (SEQ ID NO:45) was introduced by substituting the gene at the NCgl0049 gene position. The gene was introduced to a Corynebacterium pK19mobsacB using (ATCC87098). In other words, sequences of the regions upstream and downstream of the NCgl0049 gene and sequence of the cat1, sucD, and 4hbd gene regions (SEQ ID NO:45) were synthesized and ligated with pK19mobsacB at XbaI and NheI restriction enzyme sites.

The obtained plasmid was used to substitute the NCgl0049 gene with the synthesized cat1, sucD, and 4hbd genes by homologous recombination according to Schafer et al. (1994). Deletion of NCgl0049 and introduction of cat1, sucD, and 4hbd genes were verified by PCR using primer sets (SEQ ID NO: 43 and SEQ ID NO: 44). The obtained strain was named as C011.

TABLE 2

0	SEQ ID NO	Sequence	
	43	5'-ATT CGG TGA GGA ATC CGG CGG TG-3'	
5	44	5'-CTA TGA GAC AGT CGT CCT GTA CCC AT-3'	

- 1.3 Preparation of Gene-Deleted Strain for Efficient Production of 4HB
- 1.3.1 Deletion of Malate Quinone Oxidoreductase Gene
- (1) Preparation of Replacement Vector

An upstream region corresponding to first 200 bp of malate quinone oxidoreductase gene was amplified by PCR using genome DNA of CGL ATCC 13032 as a template and using primer sets (SEQ ID NO: 30 and SEQ ID NO: 31). A downstream region corresponding to last 200 bp of malate quinone oxidoreductase gene was amplified by PCR using primer sets (SEQ ID NO: 32 and SEQ ID NO: 33). Afterward, flanking regions upstream and downstream from malate quinone oxidoreductase gene was overlapped with an extended region of an oligonucleotide and linked with each other by using 16 bp In-Fusion® HD Cloning Kit (cat no. 15 639691 manufactured by Clontech). The linked regions upstream and downstream from the malate quinone oxi-

12

phosphoenolpyruvate carboxykinase product were ligated with pK19mobsacB at BamHI and SalI restriction enzyme sites. As a result, pK19mobsacB-pckG was obtained.

(2) Preparation of CGL (Δldh ΔpckG) Strain

Expression of phosphoenolpyruvate carboxykinase having an amino acid sequence of SEQ ID NO: 19 was additionally repressed. For this, a polynucleotide having a nucleic acid sequence of SEQ ID NO: 20 was deleted by homologous recombination. The obtained plasmid, the vector prepared above, pK19mobsacB (ATCC 87098), was used to delete a corresponding gene. Specifically, pckG gene was deleted by homologous recombination according to Schafer et al. (1994) by introducing the plasmid prepared above, pK19mobsacB-ΔpckG, into C011 (Δmqo) strain. Gene deletion was verified by PCR using primer sets (SEQ ID NO: 34 and SEQ ID NO: 37). The obtained strain was named as C011 (ΔmqoΔpckG).

TABLE 3

SEQ ID NO	Sequence
30	5'-CTGCAGGTCGACTCTAGAGAAGAAGTAGTCCGTCATGCCGTGAACC-3'
31	5'-TAGAAGATTATTTTTGACTGACGCGTGGGGCG-3'
32	5'-GTCAAAAATAATCTTCTAACTGCTTTCTTTAAAGCACCCG-3'
33	5'-CTCGGTACCCGGGGATCCTCTTAAAGCCTGAGATAGCGAGTTCCA-3'
34	5'-GCTCTAGAGTCATGTATTTAGGTAGGGC-3'
35	5'-ATCTGAAAGCATGCATTTGCAACGACACCAAGT-3'
36	5'-GTTGCAAATGCATGCTTTCAGATACAGAACTAG-3'
37	5'-GCTCTAGACAGTCGTTGAACTCAGGT-3'

doreductase product were ligated with pK19mobsacB at BamHI and SalI restriction enzyme sites. As a result, pK19mobsacB-Δmqo was obtained.

(2) Preparation of CGL (ΔLdh ΔMqo) Strain

In the C011 strain prepared in Example 1.2), expression of malate quinone oxidoreductase having an amino acid sequence of SEQ ID NO: 17 was additionally repressed. For this, a polynucleotide having a nucleic acid sequence of SEQ ID NO: 18 was deleted by homologous recombination. The obtained plasmid, pK19mobsacB-Δmqo, was used to delete a corresponding gene in CGL C011 by homologous recombination according to Schafer et al. (1994). Gene deletion was verified by PCR using primer sets (SEQ ID NO: 28 and SEQ ID NO: 31).

1.3.2. Deletion of Phosphoenolpyruvate Carboxykinase (1) Preparation of Replacement Vector

An upstream region corresponding to first 200 bp of 55 phosphoenolpyruvate carboxykinase gene was amplified by PCR using genome DNA of CGL ATCC 13032 as a template and using primer sets (SEQ ID NO: 34 and SEQ ID NO: 35). A downstream region corresponding to last 200 bp of phosphoenolpyruvate carboxykinase gene was amplified by 60 PCR using primer sets (SEQ ID NO: 36 and SEQ ID NO: 37). Afterward, flanking regions upstream and downstream from phosphoenolpyruvate carboxykinase gene was overlapped with an extended region of an oligonucleotide and linked with each other by using 16 bp In-Fusion® HD 65 Cloning Kit (cat no. 639691 manufactured by Clontech). The linked regions upstream and downstream from the

1.4. Preparation of Gene-Introduced Strain for Efficient Production of 4HB

1.4.1. Introduction of Pyruvate Carboxylase Gene

In an amino acid sequence of pyruvate carboxylase of the strain prepared in Examples 1.2 or 1.3, proline-458 was additionally substituted with serine. Specifically, the substitution was performed by using primers SEQ ID NO: 38 to 41.

An upstream region corresponding to first 200 bp from the proline amino acid position of pyruvate carboxylase was amplified by PCR using primer sets (SEQ ID NO: 38 and SEQ ID NO: 39). A downstream region from the proline amino acid position of pyruvate carboxylase was amplified by PCR using primer sets (SEQ ID NO: 40 and SEQ ID NO: 41). Afterward, the upstream and downstream regions substituting proline with serine in pyruvate carboxylase were overlapped with an extended region of an oligonucleotide and linked with each other by using 16 bp In-Fusion® HD Cloning Kit (cat no. 639691 manufactured by Clontech). The linked regions upstream and downstream from the phosphoenolpyruvate carboxykinase product were ligated with pK19mobsacB at BamHI and SalI restriction enzyme sites.

The obtained plasmid pK19mobsacB-mutated pyc^{P4583} was introduced to the C011 (ΔmqoΔpckG) strain by homologous recombination according to Schafer et al. (1994). Gene introduction was verified by PCR using primer sets (SEQ ID NO: 38 and SEQ ID NO:41). The obtained strain was named as C011 (ΔmqoΔpckG pyc^{P458S}).

13

TABLE 4

SEQ ID NO	Sequence
38	5'-GCTCTAGATTGAGCACACCGTGACT-3'
39	5'-CCGGATTCATTGCCGATCAC <u>T</u> C-3'
40	5'-GCTCTAGACTGTCCCACGGATCCTCAAA-3'
41	5'-CTGAAGGAGGTGCG <u>A</u> GTGA-3'

1.4.2. Introduction of α-Ketoglutarate Synthase Gene

In order to introduce α -ketoglutarate synthase gene sucA, pK19 mobsacB-sucA was prepared by introducing a synthesized polynucleotide (SEQ ID NO:46) encoding α -ketoglutarate synthase to pK19mobsacB (ATCC 87098) vector cleaved by XbaI and NheI. The obtained plasmid pK19mobsacB-sucA was introduced to the C011 (Δ mqo Δ pckG pyc P458S) strain by homologous recombination according to Schafer et al. (1994). Gene introduction was verified by PCR using primer sets (SEQ ID NO: 54 and SEQ ID NO: 55). The obtained strain was named as C098. 1.4.3. Introduction of Pyruvate Dehydrogenase

(1) Preparation of pGST1 Vector

Four PCR products were obtained by using Phusion High-Fidelity DNA Polymerase (New England Biolabs, cat.# M0530). PCR was performed by using CGL promoter screening vector pET2 (GenBank accession number: 30 AJ885178.1) as a template and using primer sequences MD-616 (SEQ ID NO: 56) and MD-618 (SEQ ID NO: 57), and using primer sequences MD-615 (SEQ ID NO: 58) and MD-617 (SEQ ID NO: 59). PCR was performed by using mammalian fluorescence protein expression vector pEGFP- 35 C1 (Clonetech) as a template and using primer sequences MD-619 (SEQ ID NO: 60) and MD-620 (SEQ ID NO: 61). PCR was performed by using E. coli cloning vector pBluescriptII SK+ as a template and using primer sequences LacZa-NR (SEQ ID NO: 62) and MD-404 (SEQ ID NO: 40 63). Each of the PCR products of 3010 bp, 854 bp, 809 bp, and 385 bp was cloned to a circular plasmid by In-Fusion EcoDry PCR Cloning Kit (Clonetech, cat.#639690) method.

The cloned vector was transformed to One Shot TOP10 Chemically Competent Cell (Invitrogen, cat.# C4040-06), 45 which was then cultured in LB culture medium including kanamycin 25 mg/L. Growing colonies were selected, and vectors were recovered from selected colonies. Then, the vector sequences were verified through full sequence analysis. The vector was named as pGSK+. To prepare a CGL 50 shuttle vector including a transcription terminator and a 3' untranslated region (UTR), a 3'UTR of CGL gltA (NCgl0795) and a rho-independent terminator of rrnB of *E. coli* rrnB were inserted to the pGSK+ vector. A 108 bp PCR fragment of gltA 3'UTR was obtained by performing PCR 55 using CGL (ATCC13032) genome DNA as a template with the primer sequences MD-627 (SEQ ID NO: 64) and MD-628 (SEQ ID NO: 65).

In addition, an rrnB transcription terminator 292 bp PCR product was obtained by performing PCR using *E. coli* 60 (MG1655) genome DNA as a template with the primer sequences MD-629 (SEQ ID NO: 66) and MD-630 (SEQ ID NO: 67). The two amplified fragments were inserted to SacI digested pGSK+ by using In-Fusion EcoDry PCR Cloning Kit (Clonetech, cat.#639690). The cloned vector was transformed to One Shot TOP10 Chemically Competent Cell (Invitrogen, cat.# C4040-06), which was then cultured in LB

14

culture medium including kanamycin 25 mg/L. Growing colonies were selected, and vectors were recovered from selected colonies. Then, the vector sequences were verified through full sequence analysis. The vector was named as pGST1.

A CGL shuttle vector wherein each gene of E. coli Pdh complex is over-expressed under NCg11929 promoter was prepared. 206 bp, 1454 bp, 2694 bp, and 1935 bp DNA fragments were obtained by performing PCR using CGL NCgl1929 promoter, Ec.lpd open reading frame (SEQ ID NO: 53) encoding E. coli dehydrolipoamide dehydrogenase (SEQ ID NO: 52) next to natural ribosome binding site, Ec.aceE open reading frame (SEQ ID NO: 49) encoding E. coli pyruvate dehydrogenase (SEQ ID NO: 48) next to natural ribosome binding site, and Ec.aceF open reading frame (SEQ ID NO: 51) encoding E. coli dihydrolipoamide acetyltransferase (SEQ ID NO:50) next to natural ribosome binding site, with primers J0180 (SEQ ID NO: 68) and MD-1081 (SEQ ID NO: 69), MD-1082 (SEQ ID NO: 70) and MD-1083 (SEQ ID NO: 71), MD-1084 (SEQ ID NO: 72) and MD-1085 (SEQ ID NO: 73), and MD-1086 (SEQ ID NO: 74) and MD-1087 (SEQ ID NO: 55), respectively.

The DNA fragments were ligated with KpnI/XbaI digested pGST1 vector using In-Fusion EcoDry PCR Cloning Kit (Clonetech, cat.#639690). The cloned vector was transformed to One Shot TOP10 Chemically Competent Cell (Invitrogen, cat.# C4040-06), which was then cultured in LB culture medium including kanamycin 25 mg/L. Vectors were recovered from the colonies. Then, the vector preparation was verified through full sequence analysis. The vector was named as MD0376. C158 strain was obtained by transforming the MD0376 vector in the form of a vector to C098 strain.

1.4.4 Introduction of Formate Dehydrogenase Gene

In order to introduce formate dehydrogenase gene, pK19 mobsacB-fdh was prepared by introducing a synthesized polynucleotide (SEQ ID NO: 47) encoding synthesized formate dehydrogenase to pK19mobsacB (ATCC 87098) vector cleaved by BamHI and EcoRI. The obtained plasmid pK19mobsacB-fdh was introduced to the C098 strain by homologous recombination according to Schafer et al. (1994). Gene introduction was verified by PCR using primer sets (SEQ ID NO: 76 and SEQ ID NO: 77). The obtained strain was named as C209.

TABLE 5

Name of strain	Genetically engineered CGL Genotype
CGL(Δldh)	ATCC13032 Δldh
C011	ATCC13032 Δldh, ΔNcgl0049, cat1, sucD, 4hbD
C098	ATCC13032 Δ ldh Δ Negl0049, cat1, sucD, 4hbD, Δ mqo, Δ pckG, pyc P458S , sucA
C158	ATCC13032 Δldh, ΔNcgl0049, cat1, sucD, 4hbD, Δmqo, ΔpckG, pyc ^{P458δ} , sucA, MD0376
C209	ATCC13032 Δldh, ΔNcgl0049, cat1, sucD, 4hbD, Δmqo, ΔpckG, pyc ^{P4588} , sucA, fdh

Example 2

Production of Various C4 Chemicals Using Transformed Strains

Each of the various CGL strains prepared in Example 1 was fermented in a fermenter at 30° C. Glucose was used as a carbon source. Fermentation was performed in anaerobic

conditions for producing various C4 chemicals. Specifically, fed-batch fermentation was performed. The culture medium included glucose 50 g/L, corn steep liquor 10 g/L, $(NH_4)_2$ SO₄ 45 g/L, urea 4.5 g/L, KH_2PO_4 0.5 g/L, $MgSO_4/7H_2O$

 $0.5~\rm g/L,\,FeSO_4/7H_2O~(10~\rm g/L)$ stock 1 mL, MnSO_4/4H_2O~(10~\rm g/L) stock 1 mL, beta-ALANIN (5~g/L) stock 1 mL, nicotinic acid (5~g/L) stock 1 mL, thiamine-HCl (5~g/L) stock 1 mL, and D-Biotin (0.3~g/L) stock 1 mL. Fermentation was performed by supplying oxygen until OD_600~80, and then by blocking oxygen supply for 100 hours.

4HB production of the strains was verified in the conditions. The basic C011 strain was verified to produce 4HB of 4.0 g/L. The additionally transformed strain C098, which was fermented in the conditions the same as those of the C011 strain, produced 28.4 g/L 4HB, which was 7.1 times the 4HB production of the C011 strain (Refer to Table 6 and FIG. 3). In addition, the C158 strain produced 40.5 g/L 4HB, which was 10.1 times and 1.4 times the 4HB production of the C011 strain and the C098 strain, respectively (Refer to Table 6 and FIG. 3). In addition, the C158 strain produced 45.7 g/L 4HB, which was 11.4 times and 1.6 times the 4HB production of the C011 strain and the C098 strain, respectively (Refer to Table 6 and FIG. 3). Under the same condition, the concentrations of succinic acid and GBL produced by the C029 strain were also measured. The C029 strain produced 12.7 g/L succinic acid and 0.3 g/L GBL, while the Δ ldh strain did not produce succinic acid or GBL.

TABLE 6

Strain	4HB Production (g/L)
∆ldh	0
C011	4
C098 C158	28.4 40.5
C209	45.7

All references, including publications, patent applications, and patents, cited herein are hereby incorporated by reference to the same extent as if each reference were individually and specifically indicated to be incorporated by reference and were set forth in its entirety herein.

16

The use of the terms "a" and "an" and "the" and "at least one" and similar referents in the context of describing the invention (especially in the context of the following claims) are to be construed to cover both the singular and the plural, unless otherwise indicated herein or clearly contradicted by context. The use of the term "at least one" followed by a list of one or more items (for example, "at least one of A and B") is to be construed to mean one item selected from the listed items (A or B) or any combination of two or more of the listed items (A and B), unless otherwise indicated herein or clearly contradicted by context. The terms "comprising," "having," "including," and "containing" are to be construed as open-ended terms (i.e., meaning "including, but not limited to,") unless otherwise noted. Recitation of ranges of values herein are merely intended to serve as a shorthand method of referring individually to each separate value falling within the range, unless otherwise indicated herein, and each separate value is incorporated into the specification as if it were individually recited herein. All methods described herein can be performed in any suitable order unless otherwise indicated herein or otherwise clearly contradicted by context. The use of any and all examples, or exemplary language (e.g., "such as") provided herein, is intended merely to better illuminate the invention and does not pose a limitation on the scope of the invention unless otherwise claimed. No language in the specification should be construed as indicating any non-claimed element as essential to the practice of the invention.

Preferred embodiments of this invention are described herein, including the best mode known to the inventors for carrying out the invention. Variations of those preferred embodiments may become apparent to those of ordinary skill in the art upon reading the foregoing description. The inventors expect skilled artisans to employ such variations as appropriate, and the inventors intend for the invention to be practiced otherwise than as specifically described herein. Accordingly, this invention includes all modifications and equivalents of the subject matter recited in the claims appended hereto as permitted by applicable law. Moreover, any combination of the above-described elements in all possible variations thereof is encompassed by the invention unless otherwise indicated herein or otherwise clearly contradicted by context.

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Val 465	Met	Val	Ile	Val	Thr 470	Glu	Gln	Gly	Val	Ala 475	Asp	Leu	Arg	Gly	Leu 480
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Lys	Ser	Ala 195	Tyr	Ser	Ser	Gly	Lys 200	Pro	Ser	Phe	Gly	Val 205	Gly	Ala	Gly
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Glu 225	Lys	Ile	Ile	Thr	Gly 230	Arg	Ala	Phe	Asp	Asn 235	Gly	Ile	Ile	CÀa	Ser 240
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Leu Arg Gln Leu Glu Pro Ser Trp Thr Gln Ile Val Phe Glu Arg Leu 35 40 45										
Asp Gly Pro Ala Gln Glu Ser Ser Pro Trp Asn Asn Ala Gly Thr 50 55 60										
Gly His Ser Ala Leu Cys Glu Leu Asn Tyr Thr Pro Glu Val Lys Gly 75 80										
Lys Val Glu Ile Ala Lys Ala Val Gly Ile Asn Glu Lys Phe Gln Val 85 90 95										
Ser Arg Gln Phe Trp Ser His Leu Val Glu Glu Gly Val Leu Ser Asp 100 105 110										
Pro Lys Glu Phe Ile Asn Pro Val Pro His Val Ser Phe Gly Gln Gly 115 120 125										
Ala Asp Gln Val Ala Tyr Ile Lys Ala Arg Tyr Glu Ala Leu Lys Asp 130 135 140										
His Pro Leu Phe Gln Gly Met Thr Tyr Ala Asp Asp Glu Ala Thr Phe 145 150 150 160										
Thr Glu Lys Leu Pro Leu Met Ala Lys Gly Arg Asp Phe Ser Asp Pro 165 170 175										
Val Ala Ile Ser Trp Ile Asp Glu Gly Thr Asp Ile Asn Tyr Gly Ala 180 185 190										
Gln Thr Lys Gln Tyr Leu Asp Ala Ala Glu Val Glu Gly Thr Glu Ile 195 200 205										
Arg Tyr Gly His Glu Val Lys Ser Ile Lys Ala Asp Gly Ala Lys Trp										
Ile Val Thr Val Lys Asn Val His Thr Gly Asp Thr Lys Thr Ile Lys										
225 230 235 240										

Ala Asn Phe Val Phe Val Gly Ala Gly Gly Tyr Ala Leu Asp Leu Leu 245 250 255
Arg Ser Ala Gly Ile Pro Gln Val Lys Gly Phe Ala Gly Phe Pro Val 260 265 270
Ser Gly Leu Trp Leu Arg Cys Thr Asn Glu Glu Leu Ile Glu Gln His 275 280 285
Ala Ala Lys Val Tyr Gly Lys Ala Ser Val Gly Ala Pro Pro Met Ser 290 295 300
Val Pro His Leu Asp Thr Arg Val Ile Glu Gly Glu Lys Gly Leu Leu 305 310 315 320
Phe Gly Pro Tyr Gly Gly Trp Thr Pro Lys Phe Leu Lys Glu Gly Ser 325 330 335
Tyr Leu Asp Leu Phe Lys Ser Ile Arg Pro Asp Asn Ile Pro Ser Tyr 340 345 350
Leu Gly Val Ala Ala Gln Glu Phe Asp Leu Thr Lys Tyr Leu Val Thr 355 360 365
Glu Val Leu Lys Asp Gln Asp Lys Arg Met Asp Ala Leu Arg Glu Tyr 370 375 380
Met Pro Glu Ala Gln Asn Gly Asp Trp Glu Thr Ile Val Ala Gly Gln 385 390 395 400
Arg Val Gln Val Ile Lys Pro Ala Gly Phe Pro Lys Phe Gly Ser Leu 405 410 415
Glu Phe Gly Thr Thr Leu Ile Asn Asn Ser Glu Gly Thr Ile Ala Gly 420 425 430
Leu Leu Gly Ala Ser Pro Gly Ala Ser Ile Ala Pro Ser Ala Met Ile 435 440 445
Glu Leu Leu Glu Arg Cys Phe Gly Asp Arg Met Ile Glu Trp Gly Asp 450 460
Lys Leu Lys Asp Met Ile Pro Ser Tyr Gly Lys Leu Ala Ser Glu 465 470 475 480
Pro Ala Leu Phe Glu Gln Gln Trp Ala Arg Thr Gln Lys Thr Leu Lys 485 490 495
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aatgaaggaa coggocacto tgototatgo gagotgaact acaccocaga ggttaagggo 240
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cgtgttcagg ttattaagcc tgcaggattc cctaagttcg gttccctgga attcggcacc	1260
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gagtggggcg acaagetgaa ggacatgate eetteetaeg gcaagaaget tgetteegag	1440
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Glu Lys Arg Pro Asn Ser Tyr Leu Ala Arg Ser Asn Pro Ser Asp Val 65 70 75 80	
Ala Arg Val Glu Ser Arg Thr Phe Ile Cys Ser Glu Lys Glu Glu Asp 85 90 95	
Ala Gly Pro Thr Asn Asn Trp Ala Pro Pro Gln Ala Met Lys Asp Glu 100 105 110	
Met Ser Lys His Tyr Ala Gly Ser Met Lys Gly Arg Thr Met Tyr Val 115 120 125	

130 135 140

150

Val Gln Leu Thr Asp Ser Glu Tyr Val Val Met Ser Met Arg Ile Met

Thr Arg Met Gly Ile Glu Ala Leu Asp Lys Ile Gly Ala Asn Gly Ser 165 \$170\$

155

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Glu	Asp	Val 195	Ala	Trp	Pro	CAa	Asn 200	Asp	Thr	Lys	Tyr	Ile 205	Thr	Gln	Phe
Pro	Glu 210	Thr	Lys	Glu	Ile	Trp 215	Ser	Tyr	Gly	Ser	Gly 220	Tyr	Gly	Gly	Asn
Ala 225	Ile	Leu	Ala	Lys	Lys 230	Cys	Tyr	Ala	Leu	Arg 235	Ile	Ala	Ser	Val	Met 240
Ala	Arg	Glu	Glu	Gly 245	Trp	Met	Ala	Glu	His 250	Met	Leu	Ile	Leu	Lys 255	Leu
Ile	Asn	Pro	Glu 260	Gly	Lys	Ala	Tyr	His 265	Ile	Ala	Ala	Ala	Phe 270	Pro	Ser
Ala	Cys	Gly 275	Lys	Thr	Asn	Leu	Ala 280	Met	Ile	Thr	Pro	Thr 285	Ile	Pro	Gly
Trp	Thr 290	Ala	Gln	Val	Val	Gly 295	Asp	Asp	Ile	Ala	Trp 300	Leu	Lys	Leu	Arg
Glu 305	Asp	Gly	Leu	Tyr	Ala 310	Val	Asn	Pro	Glu	Asn 315	Gly	Phe	Phe	Gly	Val 320
Ala	Pro	Gly	Thr	Asn 325	Tyr	Ala	Ser	Asn	Pro 330	Ile	Ala	Met	Lys	Thr 335	Met
Glu	Pro	Gly	Asn 340	Thr	Leu	Phe	Thr	Asn 345	Val	Ala	Leu	Thr	Asp 350	Asp	Gly
Asp	Ile	Trp 355	Trp	Glu	Gly	Met	Asp 360	Gly	Asp	Ala	Pro	Ala 365	His	Leu	Ile
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His 385	Pro	Asn	Ser	Arg	Tyr 390	CAa	Val	Ala	Ile	Asp 395	Gln	Ser	Pro	Ala	Ala 400
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Lys 545	Ala	Glu	Asp	Leu	Asp 550	Leu	Asp	Gly	Leu	Asp 555	Thr	Pro	Ile	Glu	Asp 560
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69 70

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His Ala

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595

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Val	Glu	Asn 35	Pro	Ala	Thr	Gly	Glu 40	Thr	Ile	Ala	Thr	Leu 45	Ala	Ser	Ala
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Thr	Tyr	Gly 115	Asn	Glu	Phe	Leu	Arg 120	Trp	Phe	Ser	Glu	Glu 125	Ala	Val	Arg
Leu	Tyr 130	Gly	Arg	Tyr	Gly	Thr 135	Thr	Pro	Glu	Gly	Asn 140	Leu	Arg	Met	Leu
Thr 145	Ala	Leu	ГÀз	Pro	Val 150	Gly	Pro	Cys	Leu	Leu 155	Ile	Thr	Pro	Trp	Asn 160
Phe	Pro	Leu	Ala	Met 165	Ala	Thr	Arg	Lys	Val 170	Ala	Pro	Ala	Ile	Ala 175	Ala
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Gln	Tyr	Phe 195	Ala	Gln	Thr	Met	Leu 200	Asp	Ala	Gly	Leu	Pro 205	Ala	Gly	Val
Leu	Asn 210	Val	Val	Ser	Gly	Ala 215	Ser	Ala	Ser	Ala	Ile 220	Ser	Asn	Pro	Ile
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Val	Gly	Gln	Gln	Leu 245	Leu	Lys	Lys	Ala	Ala 250	Asp	Lys	Val	Leu	Arg 255	Thr
Ser	Met	Glu	Leu 260	Gly	Gly	Asn	Ala	Pro 265	Phe	Ile	Val	Phe	Glu 270	Asp	Ala
Asp	Leu	Asp 275	Leu	Ala	Ile	Glu	Gly 280	Ala	Met	Gly	Ala	Lys 285	Met	Arg	Asn
Ile	Gly 290	Glu	Ala	CAa	Thr	Ala 295	Ala	Asn	Arg	Phe	Leu 300	Val	His	Glu	Ser
Val 305	Ala	Asp	Glu	Phe	Gly 310	Arg	Arg	Phe	Ala	Ala 315	Arg	Leu	Glu	Glu	Gln 320

Val Leu Gly Asn Gly Leu Asp Glu Gly Val Thr Val Gly Pro Leu Val 325 330 335 Glu Glu Lys Ala Arg Asp Ser Val Ala Ser Leu Val Asp Ala Ala Val

340 345 350

Ala Glu Gly Ala Thr Val Leu Thr Gly Gly Lys Ala Gly Thr Gly Ala 355 \$360\$

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1320

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<223> OTHER INFORMATION: Synthetic (sequences for removing LDH)
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<213> ORGANISM: Artificial Sequence

<220> FEATURE:

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<223> OTHER INFORMATION: Synthetic (sequences for removing LDH)
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<220> FEATURE:
<223> OTHER INFORMATION: Synthetic (sequences for deleting Mqo)
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<223> OTHER INFORMATION: Synthetic (sequences for deleting pckG)
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<223> OTHER INFORMATION: Synthetic (sequences for deleting pckG)
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<212> TYPE: DNA
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<220> FEATURE:
<223> OTHER INFORMATION: Synthetic (primer sequences for substituting
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<212> TYPE: DNA
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<223> OTHER INFORMATION: Synthetic (Corynebacterium glutamicum ATCC
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gaaaagaaac tcgaaggcaa cgtcatggac ttaaaccatg gtgttgtgtg ggccgattcc
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aagattatga aatccatcgt cggcgatgtc atggacagcg gattcgacgg catcttcctc
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cgaggcatcc gccgcgttgt cgaactagaa atcaccgacc acgagatgga acgcttcaag
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<210> SEO ID NO 43
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<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Synthetic (sequence for deleting gabD3)
<400> SEQUENCE: 43
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US 9,447,439 B2 -continued attcggtgag gaatccggcg gtg 23 <210> SEQ ID NO 44 <211> LENGTH: 26 <212> TYPE: DNA <213> ORGANISM: Artificial Sequence <220> FEATURE: <223> OTHER INFORMATION: Synthetic (sequence for deleting gabD3) <400> SEQUENCE: 44 ctatgagaca gtcgtcctgt acccat 26 <210> SEQ ID NO 45 <211> LENGTH: 5390 <212> TYPE: DNA <213> ORGANISM: Artificial Sequence <220> FEATURE: <223 > OTHER INFORMATION: Synthetic (polynucleotides coding succinyl-CoA: coenzyme A transferase, succinate semialdehyde dehydrogenase and 4-hydroxybutyrate dehydrogenase) <400> SEQUENCE: 45 totagaatga ctattaatgt ctccgaacta cttgccaaag tccccacggg tctactgatt 60 120 ggtgattcct gggtggaagc atccgacggc ggtactttcg atgtggaaaa cccagcgacg ggtgaaacaa tcgcaacgct cgcgtctgct acttccgagg atgcactggc tgctcttgat 180 gctgcatgcg ctgttcaggc cgagtgggct aggatgccag cgcgcgagcg ttctaatatt 240 ttacgccgcg gttttgagct cgtagcagaa cgtgcagaag agttcgccac cctcatgacc 300 360 ttggaaatgg gcaagcettt ggetgaaget egeggegaag teacetaegg eaacgaatte ctgcgctggt tctctgagga agcagttcgt ctgtatggcc gttacggaac cacaccagaa 420 ggcaacttgc ggatgctgac cgccctcaag ccagttggcc cgtgcctcct gatcacccca 480 tggaacttcc cactagcaat ggctactaga tgattttgca tctgctgcga aatctttgtt 540 teccegetaa agttgaggae aggttgaeae ggagttgaet egaegaatta tecaatgtga 600 gtaggtttgg tgcgtgagtt ggaaaaattc gccatactcg cccttgggtt ctgtcagctc 660 aagaattett gagtgaeega tgetetgatt gaeetaaetg ettgaeaeat tgeattteet 720 acaatottta gaggagacao aacatgtota aaggaatcaa gaatagocaa ttgaaaaaaa 780 agaacgtcaa ggccagtaac gttgctgaga agatcgaaga gaaggtggaa aagaccgaca 840 aggtcgttga gaaggctgct gaggtgaccg aaaagcgaat tcgaaactta aagctccagg 900 aaaaagttgt gaccgcagat gtcgcagctg acatgatcga gaatggcatg atcgtcgcaa 960

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tcct	tcad	ccg a	agcto	ggcg	eg to	gatg	gtcag	g gat	tgtg	gaac	gct	ggaa	cat (gctg	cacccg	2340
ctg	gaaa	ete «	egege	gtt	cc gt	catat	cgct	caç	ggtga	atga	acga	acgci	taa (ggcag	gtggca	2400
totacogact atatgaaact gttogotgag caggtoogta ottacgtaco ggotgacgac 246											2460					
taccgcgtac tgggtactga tggcttcggt cgttccgaca gccgtgagaa cctgcgtcac 25										2520						
cact	tega	aag 1	ttgat	gcti	c ti	atgt	cgt	g gtt	gegg	gege	tgg	gcgaa	act (ggcta	aacgt	2580
ggcg	gaaat	cg a	ataaç	gaaa	gt g	gttg	ctgad	gca	aatco	gcca	aati	tcaa	cat ·	cgato	gcagat	2640
aaag	gttaa	acc o	cgcgt	ctg	gc gt	caa										2664
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< 400)> SI	SQUEI	NCE:	50												
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Ile	Thr	Glu	Ile 20	Leu	Val	Lys	Val	Gly 25	Asp	Lys	Val	Glu	Ala 30	Glu	Gln	
Ser	Leu	Ile 35	Thr	Val	Glu	Gly	Asp 40	Lys	Ala	Ser	Met	Glu 45	Val	Pro	Ser	
Pro	Gln 50	Ala	Gly	Ile	Val	Lуз 55	Glu	Ile	Lys	Val	Ser 60	Val	Gly	Asp	Lys	
Thr 65	Gln	Thr	Gly	Ala	Leu 70	Ile	Met	Ile	Phe	Asp 75	Ser	Ala	Asp	Gly	Ala 80	
Ala	Asp	Ala	Ala	Pro 85	Ala	Gln	Ala	Glu	Glu 90	Lys	Lys	Glu	Ala	Ala 95	Pro	
Ala	Ala	Ala	Pro 100	Ala	Ala	Ala	Ala	Ala 105	Lys	Asp	Val	Asn	Val 110	Pro	Asp	
Ile	Gly	Ser 115	Asp	Glu	Val	Glu	Val 120	Thr	Glu	Ile	Leu	Val 125	Lys	Val	Gly	
Asp	Lys 130	Val	Glu	Ala	Glu	Gln 135	Ser	Leu	Ile	Thr	Val 140	Glu	Gly	Asp	ГЛа	
Ala 145	Ser	Met	Glu	Val	Pro 150	Ala	Pro	Phe	Ala	Gly 155	Thr	Val	ГÀа	Glu	Ile 160	
Lys	Val	Asn	Val	Gly 165	Asp	Lys	Val	Ser	Thr 170	Gly	Ser	Leu	Ile	Met 175	Val	
Phe	Glu	Val	Ala 180	Gly	Glu	Ala	Gly	Ala 185	Ala	Ala	Pro	Ala	Ala 190	Lys	Gln	
Glu	Ala	Ala 195	Pro	Ala	Ala	Ala	Pro 200	Ala	Pro	Ala	Ala	Gly 205	Val	Lys	Glu	

Val Asn Val Pro Asp Ile Gly Gly Asp Glu Val Glu Val Thr Glu Val 210 215 220

Met 225	Val	Lys	Val	Gly	Asp 230	Lys	Val	Ala	Ala	Glu 235	Gln	Ser	Leu	Ile	Thr 240
Val	Glu	Gly	Asp	Lys 245	Ala	Ser	Met	Glu	Val 250	Pro	Ala	Pro	Phe	Ala 255	Gly
Val	Val	Lys	Glu 260	Leu	Lys	Val	Asn	Val 265	Gly	Asp	Lys	Val	Lys 270	Thr	Gly
Ser	Leu	Ile 275	Met	Ile	Phe	Glu	Val 280	Glu	Gly	Ala	Ala	Pro 285	Ala	Ala	Ala
Pro	Ala 290	Lys	Gln	Glu	Ala	Ala 295	Ala	Pro	Ala	Pro	Ala 300	Ala	Lys	Ala	Glu
Ala 305	Pro	Ala	Ala	Ala	Pro 310	Ala	Ala	Lys	Ala	Glu 315	Gly	Lys	Ser	Glu	Phe 320
Ala	Glu	Asn	Asp	Ala 325	Tyr	Val	His	Ala	Thr 330	Pro	Leu	Ile	Arg	Arg 335	Leu
Ala	Arg	Glu	Phe 340	Gly	Val	Asn	Leu	Ala 345	Lys	Val	Lys	Gly	Thr 350	Gly	Arg
Lys	Gly	Arg 355	Ile	Leu	Arg	Glu	Asp 360	Val	Gln	Ala	Tyr	Val 365	Lys	Glu	Ala
Ile	Lys 370	Arg	Ala	Glu	Ala	Ala 375	Pro	Ala	Ala	Thr	Gly 380	Gly	Gly	Ile	Pro
Gly 385	Met	Leu	Pro	Trp	Pro 390	Lys	Val	Asp	Phe	Ser 395	Lys	Phe	Gly	Glu	Ile 400
Glu	Glu	Val	Glu	Leu 405	Gly	Arg	Ile	Gln	Lys 410	Ile	Ser	Gly	Ala	Asn 415	Leu
Ser	Arg	Asn	Trp 420	Val	Met	Ile	Pro	His 425	Val	Thr	His	Phe	Asp 430	Lys	Thr
Asp	Ile	Thr 435	Glu	Leu	Glu	Ala	Phe 440	Arg	Lys	Gln	Gln	Asn 445	Glu	Glu	Ala
Ala	Lys 450	Arg	Lys	Leu	Asp	Val 455	Lys	Ile	Thr	Pro	Val 460	Val	Phe	Ile	Met
Lys 465	Ala	Val	Ala	Ala	Ala 470	Leu	Glu	Gln	Met	Pro 475	Arg	Phe	Asn	Ser	Ser 480
Leu	Ser	Glu	Asp	Gly 485	Gln	Arg	Leu	Thr	Leu 490	Lys	ГÀз	Tyr	Ile	Asn 495	Ile
Gly	Val	Ala	Val 500	Asp	Thr	Pro	Asn	Gly 505	Leu	Val	Val	Pro	Val 510	Phe	Lys
Asp	Val	Asn 515	ГÀа	Lys	Gly	Ile	Ile 520	Glu	Leu	Ser	Arg	Glu 525	Leu	Met	Thr
Ile	Ser 530	ГÀа	ГÀа	Ala	Arg	Asp 535	Gly	Lys	Leu	Thr	Ala 540	Gly	Glu	Met	Gln
Gly 545	Gly	Cha	Phe	Thr	Ile 550	Ser	Ser	Ile	Gly	Gly 555	Leu	Gly	Thr	Thr	His 560
Phe	Ala	Pro	Ile	Val 565	Asn	Ala	Pro	Glu	Val 570	Ala	Ile	Leu	Gly	Val 575	Ser
Lys	Ser	Ala	Met 580	Glu	Pro	Val	Trp	Asn 585	Gly	Lys	Glu	Phe	Val 590	Pro	Arg
Leu	Met	Leu 595	Pro	Ile	Ser	Leu	Ser 600	Phe	Asp	His	Arg	Val 605	Ile	Asp	Gly
Ala	Asp 610	Gly	Ala	Arg	Phe	Ile 615	Thr	Ile	Ile	Asn	Asn 620	Thr	Leu	Ser	Asp
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<210> SEQ ID NO 51 <211> LENGTH: 1893

<212> TYPE: DNA <213> ORGANISM: Escherichia coli

<400> SEQUENCE: 51

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<211> LENGTH: 474

<212> TYPE: PRT

<213> ORGANISM: Escherichia coli

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Ala	Gly	Tyr	Ser 20	Ala	Ala	Phe	Arg	Сув 25	Ala	Asp	Leu	Gly	Leu 30	Glu	Thr
Val	Ile	Val 35	Glu	Arg	Tyr	Asn	Thr 40	Leu	Gly	Gly	Val	Сув 45	Leu	Asn	Val
Gly	Сув 50	Ile	Pro	Ser	Lys	Ala 55	Leu	Leu	His	Val	Ala 60	Lys	Val	Ile	Glu
Glu 65	Ala	Lys	Ala	Leu	Ala 70	Glu	His	Gly	Ile	Val 75	Phe	Gly	Glu	Pro	80 Lys
Thr	Asp	Ile	Asp	Lys 85	Ile	Arg	Thr	Trp	Lys 90	Glu	Lys	Val	Ile	Asn 95	Gln
Leu	Thr	Gly	Gly 100	Leu	Ala	Gly	Met	Ala 105	Lys	Gly	Arg	ГЛа	Val 110	Lys	Val
Val	Asn	Gly 115	Leu	Gly	rys	Phe	Thr 120	Gly	Ala	Asn	Thr	Leu 125	Glu	Val	Glu
Gly	Glu 130	Asn	Gly	Lys	Thr	Val 135	Ile	Asn	Phe	Asp	Asn 140	Ala	Ile	Ile	Ala
Ala 145	Gly	Ser	Arg	Pro	Ile 150	Gln	Leu	Pro	Phe	Ile 155	Pro	His	Glu	Asp	Pro 160
Arg	Ile	Trp	Asp	Ser 165	Thr	Asp	Ala	Leu	Glu 170	Leu	Lys	Glu	Val	Pro 175	Glu
Arg	Leu	Leu	Val 180	Met	Gly	Gly	Gly	Ile 185	Ile	Gly	Leu	Glu	Met 190	Gly	Thr
Val	Tyr	His 195	Ala	Leu	Gly	Ser	Gln 200	Ile	Asp	Val	Val	Glu 205	Met	Phe	Asp
Gln	Val 210	Ile	Pro	Ala	Ala	Asp 215	Lys	Asp	Ile	Val	Lys 220	Val	Phe	Thr	ГÀв
Arg 225	Ile	Ser	Lys	Lys	Phe 230	Asn	Leu	Met	Leu	Glu 235	Thr	Lys	Val	Thr	Ala 240
Val	Glu	Ala	ГЛа	Glu 245	Asp	Gly	Ile	Tyr	Val 250	Thr	Met	Glu	Gly	Lys 255	Lys
Ala	Pro	Ala	Glu 260	Pro	Gln	Arg	Tyr	Asp 265	Ala	Val	Leu	Val	Ala 270	Ile	Gly
Arg	Val	Pro 275	Asn	Gly	rys	Asn	Leu 280	Asp	Ala	Gly	Lys	Ala 285	Gly	Val	Glu
Val	Asp 290	Asp	Arg	Gly	Phe	Ile 295	Arg	Val	Asp	Lys	Gln 300	Leu	Arg	Thr	Asn
Val 305	Pro	His	Ile	Phe	Ala 310	Ile	Gly	Asp	Ile	Val 315	Gly	Gln	Pro	Met	Leu 320
Ala	His	Lys	Gly	Val 325	His	Glu	Gly	His	Val 330	Ala	Ala	Glu	Val	Ile 335	Ala
Gly	Lys	Lys	His 340	Tyr	Phe	Asp	Pro	Lys 345	Val	Ile	Pro	Ser	Ile 350	Ala	Tyr
Thr	Glu	Pro 355	Glu	Val	Ala	Trp	Val 360	Gly	Leu	Thr	Glu	Lys 365	Glu	Ala	ГÀз
Glu	Lys 370	Gly	Ile	Ser	Tyr	Glu 375	Thr	Ala	Thr	Phe	Pro 380	Trp	Ala	Ala	Ser
Gly 385	Arg	Ala	Ile	Ala	Ser 390	Asp	Cya	Ala	Asp	Gly 395	Met	Thr	Lys	Leu	Ile 400
Phe	Asp	Lys	Glu	Ser 405	His	Arg	Val	Ile	Gly 410	Gly	Ala	Ile	Val	Gly 415	Thr
Asn	Gly	Gly	Glu	Leu	Leu	Gly	Glu	Ile	Gly	Leu	Ala	Ile	Glu	Met	Gly

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420 425 430 Cys Asp Ala Glu Asp Ile Ala Leu Thr Ile His Ala His Pro Thr Leu 435 440 445 His Glu Ser Val Gly Leu Ala Ala Glu Val Phe Glu Gly Ser Ile Thr 455 Asp Leu Pro Asn Pro Lys Ala Lys Lys <210> SEQ ID NO 53 <211> LENGTH: 1425 <212> TYPE: DNA <213 > ORGANISM: Escherichia coli <400> SEQUENCE: 53 atgagtactg aaatcaaaac tcaggtcgtg gtacttgggg caggccccgc aggttactcc 60 getgeettee gttgegetga tttaggtetg gaaaccgtaa tegtagaacg ttacaacacc 120 cttggcggtg tttgcctgaa cgtcggctgt atcccttcta aagcactgct gcacgtagca 180 aaagttatcg aagaagccaa agcgctggct gaacacggta tcgtcttcgg cgaaccgaaa 240 300 accqatatcq acaaqattcq tacctqqaaa qaqaaaqtqa tcaatcaqct qaccqqtqqt ctqqctqqta tqqcqaaaqq ccqcaaaqtc aaaqtqqtca acqqtctqqq taaattcacc 360 ggggctaaca ccctggaagt tgaaggtgag aacggcaaaa ccgtgatcaa cttcgacaac 420 gegateattg cagegggtte tegecegate caactgeegt ttatteegea tgaagateeg 480 cgtatctggg actccactga cgcgctggaa ctgaaagaag taccagaacg cctgctggta 540 atgggtggcg gtatcatcgg tctggaaatg ggcaccgttt accacgcgct gggttcacag 600 attgacgtgg ttgaaatgtt cgaccaggtt atcccggcag ctgacaaaga catcgttaaa 660 gtcttcacca agcgtatcag caagaaattc aacctgatgc tggaaaccaa agttaccgcc 720 gttgaagcga aagaagacgg catttatgtg acgatggaag gcaaaaaagc acccgctgaa 780 ccgcagcgtt acgacgccgt gctggtagcg attggtcgtg tgccgaacgg taaaaacctc 840 gacgcaggca aagcaggcgt ggaagttgac gaccgtggtt tcatccgcgt tgacaaacag 900 ctgcgtacca acgtaccgca catctttgct atcggcgata tcgtcggtca accgatgctg 960 gcacacaaag gtgttcacga aggtcacgtt gccgctgaag ttatcgccgg taagaaacac 1020 tacttogato ogaaagttat coogtocato gootatacog aaccagaagt tgcatgggtg 1080 ggtctgactg agaaagaagc gaaagagaaa ggcatcagct atgaaaccgc caccttcccg tgggctgctt ctggtcgtgc tatcgcttcc gactgcgcag acggtatgac caagctgatt 1200 ttcgacaaag aatctcaccg tgtgatcggt ggtgcgattg tcggtactaa cggcggcgag 1260 ctgctgggtg aaatcggcct ggcaatcgaa atgggttgtg atgctgaaga catcgcactg 1320 accatccacg cgcacccgac tctgcacgag tctgtgggcc tggcggcaga agtgttcgaa 1380 1425 ggtagcatta ccgacctgcc gaacccgaaa gcgaagaaga agtag <210> SEQ ID NO 54 <211> LENGTH: 25 <212> TYPE: DNA <213> ORGANISM: Artificial Sequence <220> FEATURE: <223> OTHER INFORMATION: Synthetic (primer)

aaatggaatg gegeetagae aatee

<400> SEQUENCE: 54

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<211> LENGTH: 24
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Synthetic (primer)
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<211> LENGTH: 46
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223 > OTHER INFORMATION: Synthetic (MD-616)
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aaagtgtaaa gcctgggaac aacaagaccc atcatagttt gccccc
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<210> SEQ ID NO 57
<211> LENGTH: 36
<212> TYPE: DNA
<213 > ORGANISM: Artificial Sequence
<220> FEATURE:
<223 > OTHER INFORMATION: Synthetic (MD-618)
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gttcttctaa tcagaattgg ttaattggtt gtaaca
<210> SEQ ID NO 58
<211> LENGTH: 40
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223 > OTHER INFORMATION: Synthetic (MD-615)
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gcgtaatagc gaagagggc gtttttccat aggctccgcc
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<210> SEQ ID NO 59
<211> LENGTH: 40
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223 > OTHER INFORMATION: Synthetic (MD-617)
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gttcaatcat aacacccctt gtattactgt ttatgtaagc
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<210> SEQ ID NO 60
<211> LENGTH: 31
<212> TYPE: DNA
<213 > ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Synthetic (MD-619)
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gggtgttatg attgaacaag atggattgca c
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<210> SEQ ID NO 61
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<212> TYPE: DNA
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<223 > OTHER INFORMATION: Synthetic (MD-620)
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<220> FEATURE:
<223 > OTHER INFORMATION: Synthetic (LacZa-NR)
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cctcttcgct attacgc
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<210> SEQ ID NO 63
<211> LENGTH: 21
<212> TYPE: DNA
<213 > ORGANISM: Artificial Sequence
<220> FEATURE:
<223 > OTHER INFORMATION: Synthetic (MD-404)
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cccaggettt acaetttatg c
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<211> LENGTH: 47
<212> TYPE: DNA
<213 > ORGANISM: Artificial Sequence
<220> FEATURE:
<223 > OTHER INFORMATION: Synthetic (MD-627)
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gccaccgcgg tggagctcat ttagcggatg attctcgttc aacttcg
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<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223 > OTHER INFORMATION: Synthetic (MD-628)
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<212> TYPE: DNA
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<223 > OTHER INFORMATION: Synthetic (MD-629)
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ccgtttttgc aaataaaacg aaaggctcag tcgaaagact
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<212> TYPE: DNA
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<223> OTHER INFORMATION: Synthetic (MD-630)
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gaacaaaagc tggagctacc gtatctgtgg ggggatggct tgt
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<210> SEQ ID NO 68

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<211> LENGTH: 50
<212> TYPE: DNA
<213 > ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Synthetic (J0180)
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<210> SEQ ID NO 69
<211> LENGTH: 41
<212> TYPE: DNA
<213 > ORGANISM: Artificial Sequence
<220> FEATURE:
<223 > OTHER INFORMATION: Synthetic (MD-1081)
<400> SEQUENCE: 69
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<210> SEQ ID NO 70
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<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223 > OTHER INFORMATION: Synthetic (MD-1082)
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aaactcgaga ggaggtcatg atgagtactg aaatca
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<211> LENGTH: 36
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Synthetic (MD-1083)
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ttattcctcc tacttcttct tcgctttcgg gttcgg
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<210> SEQ ID NO 72
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<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223 > OTHER INFORMATION: Synthetic (MD-1084)
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aagaagtagg aggaataacc catgtcagaa cgtttcc
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<211> LENGTH: 26
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<213 > ORGANISM: Artificial Sequence
<220> FEATURE:
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<400> SEQUENCE: 74
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tctggcgtag gaggtaaaag aataat 26 <210> SEQ ID NO 75 <211> LENGTH: 43 <212> TYPE: DNA <213 > ORGANISM: Artificial Sequence <220> FEATURE: <223> OTHER INFORMATION: Synthetic (MD-1087) <400> SEQUENCE: 75 ggtggcggcc gctctagatt acatcaccag acggcgaatg tca 43 <210> SEQ ID NO 76 <211> LENGTH: 47 <212> TYPE: DNA <213 > ORGANISM: Artificial Sequence <220> FEATURE: <223 > OTHER INFORMATION: Synthetic (primer) <400> SEQUENCE: 76 caggtcqact ctagaggatc cgtqtcaccg atcattcqta aattgag 47 <210> SEQ ID NO 77 <211> LENGTH: 46 <212> TYPE: DNA <213 > ORGANISM: Artificial Sequence <220> FEATURE: <223> OTHER INFORMATION: Synthetic (primer) <400> SEQUENCE: 77 gtaaaacgac ggccagtgaa ttcttaggaa tcatgaaggg ggaggg 46

What is claimed is:

- 1. A genetically modified microorganism comprising
- a polynucleotide encoding exogenous α -ketoglutarate synthase, and
- a polynucleotide encoding endogenous pyruvate carboxy- 40 lase or a mutant thereof;
- wherein the genetically modified microorganism has decreased malate quinone oxidoreductase activity, decreased phosphoenolpyruvate carboxykinase activity, or a combination thereof, compared to an unmoditied microorganism of the same type,
- wherein the genetically modified microorganism produces 4-hydroxybutyrate, and
- wherein the genetically modified microorganism is a strain selected from the group consisting of lumen 50 bacteria, *Corynebacterium* genus, *Brevibacterium* genus, and *Escherichia coli*.
- 2. The genetically modified microorganism of claim 1, wherein the microorganism comprises a polynucleotide encoding succinyl-CoA:coenzyme A transferase or a mutant 55 thereof, a polynucleotide encoding coenzyme A-dependent succinate semialdehyde dehydrogenase or a mutant thereof, and a polynucleotide encoding 4-hydroxybutyrate dehydrogenase or a mutant thereof.
- 3. The genetically modified microorganism of claim 1, 60 wherein the microorganism has decreased succinate semi-aldehyde dehydrogenase activity compared to an unmodified microorganism of the same type.
- **4**. The genetically modified microorganism of claim **1**, wherein one or more of NCg10049, NCg10463, and 65 NCg12619 genes in the microorganism has an addition, substitution, or deletion mutation that eliminates succinate

- semialdehyde dehydrogenase activity, wherein the NCg10049 gene comprises the nucleic acid sequence of SEQ ID NO: 22, the NCg10463 gene comprises the nucleic acid sequence of SEQ ID NO: 23, and the NCg12619 gene comprises the nucleic acid sequence of SEQ ID NO: 24.
- **5**. The genetically modified microorganism of claim **1**, wherein the microorganism additionally comprises a polynucleotide encoding pyruvate dehydrogenase or a mutant thereof.
- **6**. The genetically modified microorganism of claim **1**, wherein the microorganism additionally comprises a gene encoding dihydrolipoyl dehydrogenase (E3), a gene encoding pyruvate dehydrogenase (E1), and a gene encoding dihydrolipoyl transacetylase (E2).
- 7. The genetically modified microorganism of claim 6, wherein the gene encoding dihydrolipoyl dehydrogenase (E3) comprises the nucleic acid sequence of SEQ ID NO: 12, the gene encoding pyruvate dehydrogenase (E1) comprises the nucleic acid sequence of SEQ ID NO: 13, and the gene dihydrolipoyl transacetylase (E2) comprises the nucleic acid sequence of SEQ ID NO: 14.
- 8. The genetically modified microorganism of claim 1, wherein the microorganism additionally comprises a polynucleotide encoding formate dehydrogenase or a mutant thereof
- **9**. The genetically modified microorganism of claim **1**, wherein the microorganism is a strain of the *Corynebacte-rium* genus.
- 10. The genetically modified microorganism of claim 9, wherein the strain of *Corynebacterium* genus is *Corynebacterium glutamicum*.

3.5

- 11. The genetically modified microorganism of claim 1, wherein the pyruvate carboxylase comprises the amino acid sequence of SEQ ID NO: 9.
- 12. The genetically modified microorganism of claim 1, wherein the mutant pyruvate carboxylase comprises the 5 amino acid sequence of SEQ ID NO: 10.
- 13. The genetically modified microorganism of claim 1, wherein the α -ketoglutarate synthase comprises the amino acid sequence of SEQ ID NO: 7.
- 14. The genetically modified microorganism of claim 2, wherein the succinyl-CoA:coenzyme A transferase comprises the amino acid sequence of SEQ ID NO: 1, the CoA-dependent succinate semialdehyde dehydrogenase comprises the amino acid sequence of SEQ ID NO: 3, and the 4-hydroxybutyrate dehydrogenase comprises the amino acid sequence of SEQ ID NO: 5.
- 15. The genetically modified microorganism of claim 8, wherein the formate dehydrogenase comprises the amino acid sequence of SEQ ID NO: 15.

122

16. A method of producing a C4-chemical comprising: culturing the genetically modified microorganism of claim **1** in a cell culture medium, whereby the microorganism produces a C4-chemical; and

recovering the C4-chemical from the cell culture medium.

- 17. The method of claim 16, wherein the C4-chemical is 4-hydroxybutyrate.
- **18**. The method of claim **16**, wherein the genetically modified microorganism additionally comprises a polynucleotide encoding pyruvate dehydrogenase or a mutant thereof and a polynucleotide encoding formate dehydrogenase or a mutant thereof.
- 19. The method of claim 18, wherein the C4-chemical is selected from the group consisting of succinic acid, 4-hydroxybutyrate, and gamma butyrolactone.

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